

ORIGINAL ARTICLE

Comparison of different protocols for ^{13}C -urea breath test for the diagnosis of *Helicobacter pylori* infection in healthy volunteers

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Abstract

Objective. The ^{13}C -urea breath test (^{13}C -UBT) is the most accurate non-invasive method for diagnosis of *Helicobacter pylori* infection. However, several methodological issues have not been resolved yet. The aim of this study was to test different protocols of ^{13}C -UBT to find the optimal test drink and sampling interval. **Material and methods.** ^{13}C -UBT was performed at 3-day intervals in 27 healthy volunteers using citric acid (test A), orange juice (B) and still water (C) as test drinks. Breath samples were collected from time 5 to 60 min. A total number of 2106 breath samples were analysed by isotope ratio mass spectrometry (cut-off value 3.5). **Results.** Differences in delta values were greater than would be expected by chance (A versus B and A versus C at times 20, 25, 30, 35 and 40 min, $p < 0.05$, Dunnett's method). There were no grey zone- or false-negative results among *H. pylori*-positive persons in test A at any time, but some were found in tests B and C. Optimal intervals for breath sampling are at times 20 or 25 min after ^{13}C -urea ingestion. **Conclusions.** Citric acid solution as a test drink and 20- or 25-min breath sampling intervals are optimal for the ^{13}C -UBT in healthy volunteers.

Key Words: Citric acid, *Helicobacter pylori*, isotope ratio mass spectrometry, test drink, ^{13}C -urea breath test

Introduction

The ^{13}C -urea breath test (^{13}C -UBT) is the most accurate non-invasive method to diagnose *Helicobacter pylori* infection [1]. The test is based on the gastric urease activity of *H. pylori* which splits non-radioactive stable ^{13}C -labelled urea ingested by investigated persons into

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NH_4^+ and ^{13}C -labelled HCO_3^- , which is expired as $^{13}\text{CO}_2$ in the exhaled breath. Expired ^{13}C -labelled carbon dioxide is measured as a $^{13}\text{CO}_2/^{12}\text{CO}_2$ ratio and results are expressed as the excess delta (difference between δt_i and δt_0).

Although the original ^{13}C -UBT was published as early as in 1987 [2], the test has not been unified yet. A European standard protocol [3] and US protocols [4,5] were proposed but not generally accepted. There are more than 20 commercial producers of diagnostic kits of ^{13}C -UBT with different settings. Several protocol modifications have been published. Last but not least, breath samples of ^{13}C -UBT can be measured by different methods, by means of isotope ratio mass spectrometry (gold standard), isotope-selective non-dispersive infrared spectrometry or by optogalvanic laser spectroscopy. Several issues have not yet been definitely resolved, e.g. different amount of ^{13}C -urea used, type of test drink (meal), optimal interval(s) for collecting breath samples or cut-off value. The aim of our study was to test different protocols of ^{13}C -UBT to find the optimal test drink and sampling interval.

Material and methods

Subjects

Twenty-seven healthy volunteers entered the study (12 M, 15 F, age 22–52 years, median 29). The ^{13}C -UBT was performed at 3-day intervals in all persons using three different protocols. Written informed consent for participation in the study was obtained from all individuals.

Urea breath test

Urea breath tests were performed in the morning after overnight fasting. Two baseline breath samples were taken before the test, followed by test drink ingestion: 150 ml still water with 3 g citric acid (test A) or 150 ml natural orange juice (test B) or 150 ml still water only (test C). After 5 min (at time 0) all persons ingested 100 mg ^{13}C -urea dissolved in 50 ml still water with 1 g citric acid (test A) or 50 ml natural orange juice (test B) or 50 ml still water only (test C).

Breath samples were collected in duplicate using a straw in 20-ml vacutainers at 5-min intervals from time 5 to 60 min, with the subjects lying on their left side.

A total of 2106 breath samples were analysed by means of isotope ratio mass spectrometry (AP 2003, Analytical Precision, UK). Cut-off was set at 3.5, based on our previous study of 1476 analyses [6].

Statistics

Data were analysed statistically with the Mann-Whitney rank sum test, the Kruskal-Wallis analysis of variance on ranks (ANOVA on ranks) and Dunnett's multiple comparisons using statistical software (SigmaStat, Jandel Corporation, Germany).

Results

Six persons (22%) were *H. pylori* positive and 21 subjects (78%) were *H. pylori* negative, clearly distinguished in all three tests (see Figures 1–3). Delta values were significantly

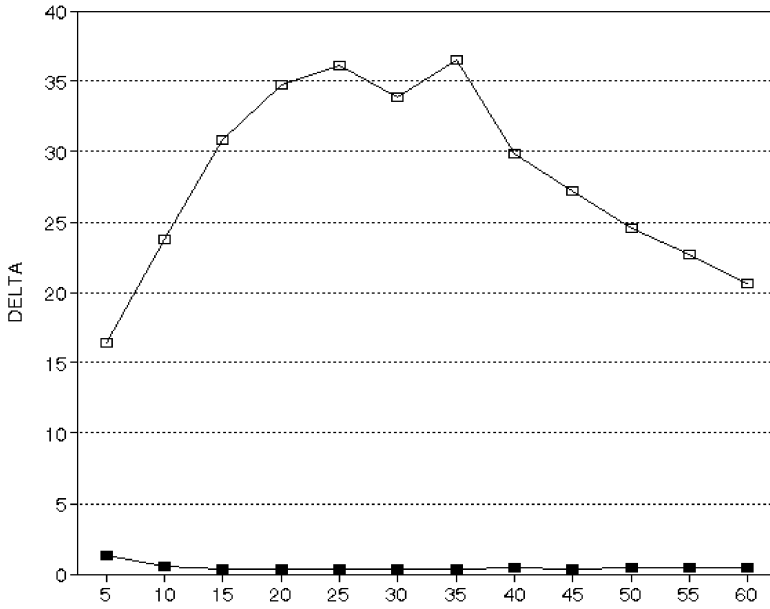


Figure 1. Test A. Average delta values of ¹³C-urea breath test at 5-min intervals (from 5 to 60 min). Citric acid solution was used as a test drink. ■: *H. pylori*-negative persons (21 subjects); □: *H. pylori*-positive persons (6 subjects).

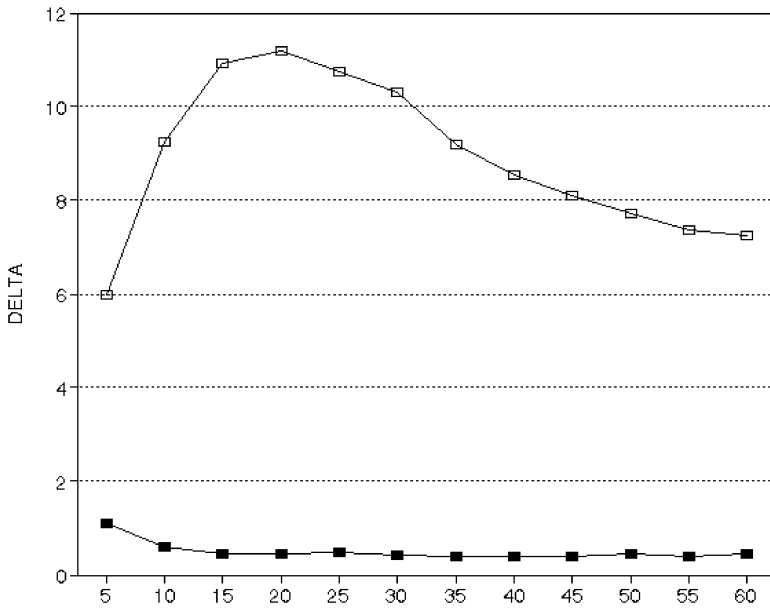


Figure 2. Test B. Average delta values of ¹³C-urea breath test at 5-min intervals (from 5 to 60 min). Natural orange juice was used as a test drink. ■: *H. pylori*-negative persons (21 subjects); □: *H. pylori*-positive persons (6 subjects).

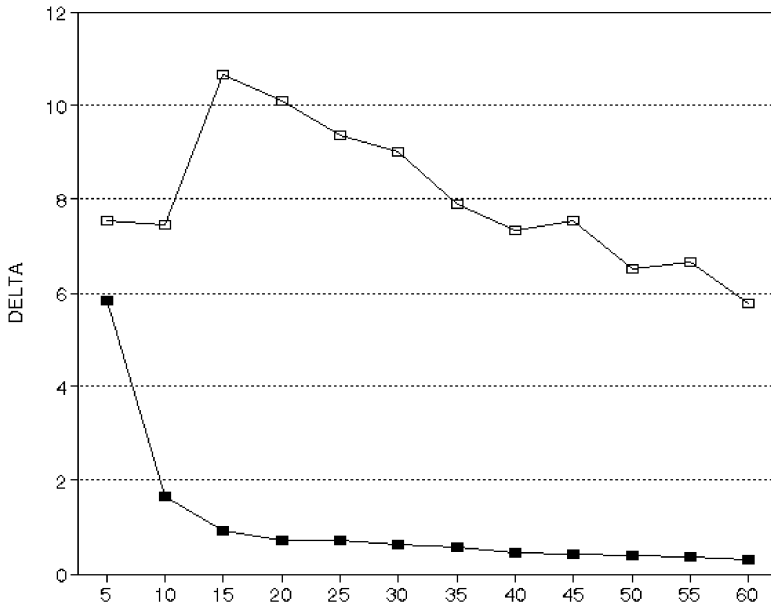


Figure 3. Test C. Average delta values of ¹³C-urea breath test at 5-min intervals (from 5 to 60 min). Still water was used as a test drink. ■: *H. pylori*-negative persons (21 subjects); □: *H. pylori*-positive persons (6 subjects).

higher in test A (citric acid) compared with test B (orange juice) and test C (water) (Table I). Differences in the median values among the group were greater than would be expected by chance (A versus B and A versus C at 20, 25, 30, 35 and 40 min, $p < 0.05$, Dunnett's method). There were no grey zone- or false-positive results among *H. pylori*-negative persons in any test at any time (specificity 100%). There were no grey zone- or false-negative results among *H. pylori*-positive persons in the test A (using citric acid as a test drink) at any time, but some were found in tests B and C (see Table II). Using citric

Table I. Delta values of ¹³C-UBT in *H. pylori*-positive persons given as median and interquartile range. Different test drinks were used: citric acid (test A), orange juice (test B) or still water (test C).

Delta	Test A	Test B	Test C	Significance
Time 20	19.618	6.510	6.774	A versus B: $p=0.004$
	12.956–52.248	5.911–15.352	3.507–7.326	A versus C: $p=0.002$ B versus C: NS
Time 25	21.041	7.587	5.819	A versus B: $p=0.003$
	13.791–50.933	6.266–15.353	3.576–8.646	A versus C: $p=0.001$ B versus C: NS
Time 30	19.618	6.510	6.774	A versus B: $p=0.004$
	12.956–52.248	5.911–15.352	3.507–7.326	A versus C: $p=0.002$ B versus C: NS
Time 35	22.371	5.721	4.992	A versus B: $p=0.002$
	13.289–52.878	5.153–12.479	2.822–6.912	A versus C: $p=0.002$ B versus C: NS
Time 40	16.573	5.726	4.495	A versus B: $p=0.002$
	11.666–50.228	4.635–11.326	2.448–7.001	A versus C: $p=0.001$ B versus C: NS

Abbreviations: ¹³C-UBT=¹³C-urea breath test; NS=not significant.

Table II. Grey zone and false-negative results in *H. pylori*-positive persons in test B (orange juice) and C (water). Two breath samples were analysed in each person at each time. Delta values are given as mean ± SD.

Time	Test C grey zone results (3.3–3.7)	Test B grey zone results (3.3–3.7)	Test C false-negative results (<3.3)	Test B false-negative results (<3.3)
5	3.599 <i>n</i> =1	None	None	2.516 <i>n</i> =1
10	None	None	None	None
15	None	None	None	None
20	None	None	None	None
25	3.577 <i>n</i> =1	None	3.039 <i>n</i> =1	None
30	None	None	3.051 ± 0.307 <i>n</i> =2	None
35	3.531 <i>n</i> =1	None	2.515 ± 0.383 <i>n</i> =2	None
40	None	3.693 <i>n</i> =1	2.435 ± 0.452 <i>n</i> =3	None
45	None	3.557 <i>n</i> =1	2.141 ± 0.570 <i>n</i> =3	None
50	None	3.650 <i>n</i> =1	1.815 ± 0.213 <i>n</i> =3	None
55	None	3.650 <i>n</i> =1	1.614 ± 0.119 <i>n</i> =3	2.728 <i>n</i> =1
60	None	None	1.396 ± 0.229 <i>n</i> =3	2.245 ± 0.263 <i>n</i> =2

acid solution as a test drink, the optimal time intervals for breath sampling are at 20 or 25 min after ¹³C-urea ingestion.

Discussion

The ¹³C-UBT is a simple, non-invasive and reliable test for the diagnosis of *H. pylori* infection. In our previous studies using citric acid solution as a test drink, we found excellent reproducibility of both analytical [6] and clinical [7] results of ¹³C-UBT. In this study we used citric acid solution, still water or orange juice as test drinks in ¹³C-UBT performed three times in the same group subjects at 3-day intervals. The best results were reached with citric acid; optimal intervals for breath sampling were at 20 or 25 min after ¹³C-urea ingestion. In agreement with other studies [8], citric acid was superior to both orange juice and still water.

Since the first description of ¹³C-UBT by Graham et al. [2] several modifications have been published to simplify and optimize the test, including the amount of ¹³C-urea used, type of test meal, number of samples and collecting time. Comparison of different protocols and the analytical methods used must always be taken into consideration, too (mass spectrometry or infrared spectrometry or optogalvanic laser spectroscopy).

Originally, different test nutrient meals (solid, semi-solid or liquid) were used to slow gastric emptying and to maximize distribution of the substrate within the stomach. Recommended test doses of ¹³C-urea have successively decreased from an initial 5 mg/kg body-weight [2] to 75 mg [9–11], 50 mg [12–15], 38 mg [16] or even 25 mg [17], and time of the breath test has shortened from an initial 180 min, collecting samples every 10 min [2], to baseline one and another one test breath sample at 20 min [5,14,18–20], 15

[15,21,22], 10 [12,23] or even 5 min [17]. Different test drinks (instead of test meal) were introduced, e.g. citric acid [5,8–12,24–28], orange juice [29], apple juice [30], fresh milk [31], water [20,32], or even no test meal/drink [5,33]. Even fasting before a ^{13}C -UBT may not be necessary according to some investigators [34,35].

Several papers have been published suggesting that citric acid solution is the optimal test drink [5,8–12,24–28]. Intra-gastric urease activity is pH-dependent, low at neutral pH, but as the external pH decreases to 6.5–5.5 there is a 10–20-fold increase in urease activity which remains high through approximately pH 2.5 [36,37]. Proton pump inhibitors (PPIs) may cause false-negative results of ^{13}C -UBT by unknown mechanism [38–42]. PPI-induced false negative results can be associated with a direct anti-*H. pylori* effect of the PPI (inhibition of the viability and growth of *H. pylori* resulting in decreased bacterial load) or associated with increased intra-gastric pH. According to some investigators [43], ^{13}C -UBT can be and according to others [44] cannot be prevented by acidification of the stomach. H_2 -receptor antagonists differ from PPIs, as high intra-gastric pH may cause a reduction in urease activity, unrelated to a reduced bacterial load, and can be reversed by citric acid [44].

Citric acid slows gastric emptying in a dose-dependent manner [11,45] and some investigators have hypothesized that prolonged contact of ^{13}C -urea with bacteria results in increased amounts of hydrolyzed urea [23,24]. Later papers showed that *H. pylori* urease activity enhanced by citric acid is unrelated to rate of gastric emptying [30,45]. Activation of the cytoplasmic urease by urea entry into the cytoplasm of *H. pylori* may explain the increased $^{13}\text{CO}_2$ excretion when an acid test meal is given rather than delayed gastric emptying [30].

To summarize our results, citric acid solution as a test drink and 20- or 25-min breath sampling intervals are optimal for the ^{13}C -UBT in healthy volunteers.

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